# **Pyruvic Acid Colorimetric Assay Kit**

Catalog No: E-BC-K130-M

Method: Colorimetric method

Specification: 96T (Can detect 80 samples without duplication)

Measuring instrument: Microplate reader

Sensitivity: 0.003 µmol/mL

Detection range: 0.003-2.0 µmol/mL

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

# **General information**

#### ▲ Intended use

This kit can be used to measure pyruvic acid content in serum, plasma and tissue samples.

### **▲** Background

Pyruvic acid is the simplest of the alpha-keto acids, with a carboxylic acid and a ketone functional group. Pyruvic acid can be made from glucose through glycolysis, converted back to carbohydrates (such as glucose) via gluconeogenesis, or to fatty acids through a reaction with acetyl-CoA. It can also be used to construct the amino acid alanine and can be converted into ethanol or lactic acid via fermentation.

Pyruvic acid supplies energy to cells through the citric acid cycle (also known as the Krebs cycle) when oxygen is present (aerobic respiration), and alternatively ferments to produce lactate when oxygen is lacking (lactic acid fermentation).

# ▲ Detection principle

Pyruvic acid can react with chromogenic agent and the reaction product is reddish brown in alkaline solution. The depth of color is directly proportional to the pyruvate content. The pyruvate content can be calculated by measuring the OD value at 505 nm

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# ▲ Kit components & storage

Item	Component	Specification	Storage	
Reagent 1	Clarificant	1.2 mL × 1 vial	2-8°C , 6 months	
Reagent 2	Chromogenic Agent	6 mL × 1 vial	2-8°C , 6 months, shading light	
Reagent 3	Alkali Reagent	20 mL × 1 vial	2-8°C , 6 months	
Reagent 4	2 µmol/mL Sodium Pyruvate Standard	1.6 mL × 1 vial	2-8°C , 6 months	
	Microplate	96 wells	No requirement	
	Plate Sealer	2 pieces		

NNote: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

# ▲ Materials prepared by users



# **Instruments**

Microplate reader (480-520 nm), Micropipettor, Centrifuge, Incubator, Vortex mixer

# Consumptive material

Tips (10  $\mu$ L, 200  $\mu$ L, 1000  $\mu$ L)

# Reagents:

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

# ▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

#### **▲ Precautions**

Before the experiment, please read the instructions carefully, and wear gloves and work clothes



# **Pre-assay preparation**

# ▲ Sample preparation

The samples should be prepared as conventional methods. Also please refer to appendix II.

# ▲ Dilution of sample

It is recommended to take 2~3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (0.003-2.0 µmol/mL).

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor	
Human serum	1	
Mouse serum	1	
Mouse plasma	1	
10% Mouse liver tissue homogenate	1	
10% Rat kidney tissue homogenate	1	
10% Rat heart tissue homogenate	1	

Note: The diluent is normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

Assay protocol				
Ambient temperature	25-30℃			
Optimum detection wavelength	505 nm			

# Instructions for the use of transferpettor:

- (1) Equilibrate the pipette tip in that reagent before pipetting each reagent.
- (2) Don't add the liquid outside the tips into the reaction system when pipetting each reagent.

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# Assay protocol

# ▲ Plate set up

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Α	Α	S1	S9	S17	S25	S33	S41	S49	S57	S65	S73
В	В	В	S2	S10	S18	S26	S34	S42	S50	S58	S66	S74
С	С	С	S3	S11	S19	S27	S35	S43	S51	S59	S67	S75
D	D	D	S4	S12	S20	S28	S36	S44	S52	S60	S68	S76
Е	Е	Е	S5	S13	S21	S29	S37	S45	S53	S61	S69	S77
F	F	F	S6	S14	S22	S30	S38	S46	S54	S62	S70	S78
G	G	G	S7	S15	S23	S31	S39	S47	S55	S63	S71	S79
Н	Н	Н	S8	S16	S24	S32	S40	S48	S56	S64	S72	S80

[Note]: A-H, standard wells; S1-S80, sample wells.

# **▲** Operating steps

#### 1. The preparation of standard curve

Dilute 2 µmol/mL sodium pyruvate standard with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 0.1, 0.2, 0.6, 0.8, 1.2, 1.6, 2 µmol/mL.

#### 2. The measurement of samples

- a. For serum (plasma) sample
  - Standard well: Add 15 µL of standard solution with different concentrations and 50 µL of reagent 2 to the corresponding wells.
    - Sample well: Add 15  $\mu L$  of sample and 50  $\mu L$  of reagent 2 to the corresponding wells.
  - 2) Mix fully with microplate reader for 10 sec, then incubate at 37°C for 10 min.
  - Add 150 μL of reagent 3 into each well. Mix fully with microplate reader for 10 sec, then incubate at room temperature for 5 min.
  - 4) Measure the OD value of each well at 505 nm with microplate reader.
- b. For tissue sample
  - Standard well: Add 15 µL of standard solution with different concentrations to the corresponding wells.
    - Sample well: Add 15 µL of sample to the corresponding wells.
  - 2) Add 10 µL of reagent 1 and 50 µL of reagent 2 to each well.
  - 3) Mix fully with microplate reader for 10 sec, then incubate at 37°C for 10 min.
  - 4) Add 150 µL of reagent 3 into each well. Mix fully with microplate reader for 10 sec, then incubate at room temperature for 5 min.
  - 5) Measure the OD value of each well at 505 nm with microplate reader.



# **▲** Operation table

# 1. For serum (plasma) sample

	Standard well	Sample well			
Standard solution with different concentrations (µL)	15				
Sample (µL)		15			
Reagent 2 (µL)	50	50			
Mix fully with microplate reader for 10 sec, then incubate at 37℃ for 10 min.					
Reagent 3 (µL)	150	150			
Mix fully with microplate reader for 10 sec, then incubate at room temperature					

Mix fully with microplate reader for 10 sec, then incubate at room temperature for 5 min. Measure the OD value of each well at 505 nm with microplate reader.

# 2. For tissue sample

	Standard well	Sample well		
Standard solution with different	15			
concentrations (µL)	15			
Sample (µL)		15		
Reagent 1 (µL)	10	10		
Reagent 2 (µL)	50	50		
Mix fully with microplate reader for 10 sec, then incubate at 37°C for 10 min.				
Reagent 3 (µL)	150	150		
Mix fully with microplate reader for 10 sec, then incubate at room temperature for				

Mix fully with microplate reader for 10 sec, then incubate at room temperature for 5 min. Measure the OD value of each well at 505 nm with microplate reader.

#### **▲** Calculation

Plot the standard curve by using OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the OD value of sample. The standard curve is: y=ax+b.

1.Serum/plasma samples:

Pyruvic Acid (
$$\mu$$
mol/mL) = ( $\Delta A_{505}$  - b)  $\div$  a × f

2. Tissue samples:

Pyruvic Acid (
$$\mu$$
mol/mgprot) = ( $\Delta A_{505}$  - b) ÷ a × f ÷ C<sub>pr</sub>

#### Note:

y: OD<sub>Standard</sub> - OD<sub>Blank</sub>.

x: The concentration of standard.

a: The slope of standard curve.

b: The intercept of standard curve.

f: Dilution factor of sample before tested.

 $\Delta A_{505}$ :  $OD_{Sample} - OD_{Blank}$ .

C<sub>pr</sub>: Concentration of protein in sample, mgprot/mL.

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# **▲ Notes**

- 1. This kit is for research use only.
- 2. Instructions should be followed strictly, changes of operation may result in unreliable results.
- 3. The validity of kit is 6 months.
- 4. Do not use components from different batches of kit.

# **Appendix I Performance characteristics**

Appendix I Performance characteristics						
Detection range	Average intra-assay CV (%)	2.3				
Sensitivity	0.003 µmol/mL	Average inter-assay CV (%)	3.6			
Average recovery rate (%)	95					

# ▲ Example analysis

Take 15 µL of human serum sample and carry the assay according to the operation table. The results are as follows:

Standard curve: y = 0.4983 x + 0.0123, the average OD value of the sample is 0.269, the average OD value of the blank is 0.047, and the calculation result is:

Pyruvic Acid 
$$= \frac{0.269 - 0.047 - 0.0123}{0.4983} = 0.42 \ \mu mol/mL$$



# **Appendix II Sample preparation**

The following sample pretreatment methods are for reference only.

#### Serum

Collect fresh blood and stand at 25°C for 30 min to clot the blood. Then centrifuge at 2000 g for 15 min at 4°C. Take the serum (which is the upper light yellow clarified liquid layer) to preserve it on ice for detection. If not detected on the same day, the serum can be stored at -80°C for a month.

#### ▲ Plasma

Take fresh blood into the tube which has anticoagulant, centrifuge at 700-1000 g for 10 min at 4°C . Take the plasma (which is the upper light yellow clarified liquid layer, don't take white blood cells and platelets in the middle layer) to preserve it on ice for detection. If not detected on the same day, the plasma can be stored at -80°C for a month.

# ▲ Tissue sample

Take 0.02-1g fresh tissue to wash with PBS (0.01 M, pH 7.4) at 2-8°C . Absorb the water with filter paper and weigh. Homogenize at the ratio of the volume of homogenized medium (2-8 $^{\circ}$ C) (mL): the weight of the tissue (g) =9:1, then centrifuge the tissue homogenate for 10 min at 10000 g at 4°C. Take the supernatant to preserve it on ice for detection. Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M). If not detected on the same day, the tissue sample (without homogenization) can be stored at -80°C for a month.

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#### Note:

- 1. Homogenized medium: Normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).
- 2. Homogenized method:
- (1) Hand-operated: Weigh the tissue and mince to small pieces (1mm<sup>3</sup>), then put the tissues pieces to glass homogenized tube. Add homogenized medium into homogenized tube, place the tube into the ice bath with left hand, and insert the glass tamping rod vertically into the homogenized tube with the right hand to grind up and down for 6-8 min.
  - Or put the tissue into the mortar, and add liquid nitrogen to grind fully. Then add the homogenized medium to homogenize.
- (2) Mechanical homogenate: Weigh the tissue to EP tube, add the homogenized medium to homogenize the tissue with homogenizer instrument (60 Hz. 90s) in the ice bath. (For samples of skin, muscle and plant tissue, the time of homogenization can be properly prolonged.)

### Notes for sample

- 1. Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
- 2. If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.
- 3. If a lysis buffer is used to prepare tissue homogenates, there is a possibility of causing a deviation due to the introduced chemical substance.