

Iron Colorimetric Assay Kit

Catalog No: E-BC-K139-M

Method: Colorimetric method

Specification: 96T (Can detect 80 samples without duplication)

Measuring instrument: Microplate reader

Sensitivity: 0.08 mg/L

Detection range: 0.29-10 mg/L

- ▶ Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

General information

▲ Intended use

This kit can measure Iron content in serum, tissue samples.

▲ Background

Iron is an essential biological element for most organisms, from bacteria to mammals. It is usually stored in metalloprotein centers, heme complexes, and oxygen carrier proteins. Most of the iron stored in the body is ferritin, but as iron is overloaded, the proportion of iron sulfur protein will increase. Iron is essential for many metabolic processes, including oxygen transport, DNA synthesis, and electron transport.

▲ Detection principle

Under the action of acidic solution and reductant, ferric ions can be separated from transferrin in serum, and reduced into ferrous ions (Fe^{2+}). The latter then bind to bipyridine and form pink complexes. The concentration of iron can be calculated by measuring the OD value at 520 nm indirectly.

▲ Kit components & storage

Item	Component	Specification	Storage
Reagent 1	10 mg/L Iron Standard	1 mL × 2 vials	2-8°C , 6 months
Reagent 2	Chromogenic Agent A	Powder × 2 vials	2-8°C , 6 months, shading light
Reagent 3	Chromogenic Agent B	Powder × 2 vials	2-8°C , 6 months, shading light
Reagent 4	Chromogenic Agent C	20 mL × 2 vials	2-8°C , 6 months
	Microplate	96 wells	No requirement
	Plate Sealer	2 pieces	

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

▲ Materials prepared by users

Instruments

Microplate reader (510-530 nm), Micropipettor, Centrifuge, Water bath, Incubator, Vortex mixer

Consumptive material

Tips (10 μ L, 200 μ L, 1000 μ L), EP tubes (1.5 mL, 2 mL)

Reagents:

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

▲ Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes.

▲ The key points of the assay

1. The supernatant after centrifugation must be clarified, and if there is turbidity, it must be centrifuged again.
2. During the experiment, the experimental vessel must be clean to avoid iron contamination which may affect the result of the experiment.
3. Iron chromogenic agent should be prepared in advance because the reagent 2 powder and reagent 3 powder are difficult to dissolved.

Pre-assay preparation

▲ Reagent preparation

Preparation of iron chromogenic agent :

Dissolve 1 vial of reagent 2 and 1 vial of reagent 3 with 20 mL of reagent 4. The prepared solution can be stored at 2-8°C for 1 month with shading light.

▲ Sample preparation

The samples should be prepared as conventional methods. Also please refer to appendix II.

Sample requirements

Metal chelating agents such as EDTA and citrate cannot be added to the sample.

▲ Dilution of sample

It is recommended to take 2~3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (0.29-10 mg/L).

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
Human serum	1
Mouse serum	1
10% Mouse liver tissue homogenate	1
10% Rat kidney tissue homogenate	1

Note:The diluent is normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

Assay protocol	
Ambient temperature	25-30°C
Optimum detection wavelength	520 nm

Instructions for the use of transferpettor:

- (1) Equilibrate the pipette tip in that reagent before pipetting each reagent.
- (2) Don't add the liquid outside the tips into the reaction system when pipetting each reagent.

Assay protocol

▲ Plate set up

	1	2	3	4	5	6	7	8	9	10	11	12
A	A	A	S1	S9	S17	S25	S33	S41	S49	S57	S65	S73
B	B	B	S2	S10	S18	S26	S34	S42	S50	S58	S66	S74
C	C	C	S3	S11	S19	S27	S35	S43	S51	S59	S67	S75
D	D	D	S4	S12	S20	S28	S36	S44	S52	S60	S68	S76
E	E	E	S5	S13	S21	S29	S37	S45	S53	S61	S69	S77
F	F	F	S6	S14	S22	S30	S38	S46	S54	S62	S70	S78
G	G	G	S7	S15	S23	S31	S39	S47	S55	S63	S71	S79
H	H	H	S8	S16	S24	S32	S40	S48	S56	S64	S72	S80

[Note] : A - H, standard wells; S1 - S80, sample wells.

▲ Operating steps

1. The preparation of standard curve

Dilute 10 mg/L iron standard stock solution with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 1, 2, 4, 5, 6, 8, 10 mg/L.

2. The measurement of samples

- 1) **Standard tube:** Add 75 μL of standard solution with different concentrations to the tubes.
Sample tube: Add 75 μL of sample to the tubes.
- 2) Add 300 μL of iron chromogenic agent, mix fully with vortex mixer.
- 3) Incubate the tubes in 100°C water bath for 5 min.
- 4) Cool the tubes with running water, centrifuge the tubes at 3000 g for 10 min.
- 5) Take 200 μL of supernatant to the microplate. Measure the OD value of each well with microplate reader at 520 nm.

▲ Operation table

	Standard tube	Sample tube
Standard solution with different concentrations (μL)	75	
Sample (μL)		75
Iron chromogenic agent (μL)	300	300
Mix fully with vortex mixer, incubate the tubes in 100°C water bath for 5 min. Cool the tubes with running water, centrifuge the tubes at 3000 g for 10 min. Take 200 μL of supernatant to the microplate. Measure the OD value of each well with microplate reader at 520 nm.		

▲ Calculation

Plot the standard curve by using OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the OD value of sample. The standard curve is: $y = ax + b$.

1. Serum/plasma samples:

$$\text{Fe content (mg/L)} = (\Delta A_{520} - b) \div a \times f$$

2. Tissue samples:

$$\text{Fe content (mg/gprot)} = (\Delta A_{520} - b) \div a \times f \div C_{pr}$$

Note:

y: $OD_{\text{Standard}} - OD_{\text{Blank}}$

x: The concentration of standard.

a: The slope of standard curve.

b: The intercept of standard curve.

f: Dilution factor of sample before tested.

ΔA_{520} : $OD_{\text{Sample}} - OD_{\text{Blank}}$

C_{pr} : Concentration of protein in sample, gprot/L.

▲ Notes

1. This kit is for research use only.
2. Instructions should be followed strictly, changes of operation may result in unreliable results.
3. The validity of kit is 6 months.
4. Do not use components from different batches of kit.

Appendix I Performance characteristics

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Detection range	0.29-10 mg/L	Average intra-assay CV (%)	1.2
Sensitivity	0.08 mg/L	Average inter-assay CV (%)	2.8
Average recovery rate (%)	96		

▲ Example analysis

Take 75 μ L of human serum sample and carry the assay according to the operation table. The results are as follows:

Standard curve: $y = 0.01427x + 0.0008$, the average OD value of the sample is 0.078, the average OD value of the blank is 0.038, and the calculation result is:

$$\text{Fe content (mg/L)} = \frac{0.078 - 0.038 - 0.0008}{0.01427} = 2.75 \text{ mg/L}$$

Appendix II Sample preparation

The following sample pretreatment methods are for reference only.

▲ Serum

Collect fresh blood and stand at 25°C for 30 min to clot the blood. Then centrifuge at 2000 g for 15 min at 4°C . Take the serum (which is the upper light yellow clarified liquid layer) to preserve it on ice for detection. If not detected on the same day, the serum can be stored at -80°C for a month.

▲ Tissue sample

Take 0.1-1g fresh tissue to wash with PBS (0.01 M, pH 7.4) at 2-8°C . Absorb the water with filter paper and weigh. Homogenize at the ratio of the volume of homogenized medium (2-8°C) (mL): the weight of the tissue (g) =9:1, then centrifuge the tissue homogenate for 10 min at 10000 g at 4°C . Take the supernatant to preserve it on ice for detection. Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M). If not detected on the same day, the tissue sample (without homogenization) can be stored at -80°C for a month.

Note:

1. Homogenized medium: Normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

2. Homogenized method:

(1) Hand-operated: Weigh the tissue and mince to small pieces (1 mm^3), then put the tissues pieces to glass homogenized tube. Add homogenized medium into homogenized tube, place the tube into the ice bath with left hand, and insert the glass tamping rod vertically into the homogenized tube with the right hand to grind up and down for 6-8 min.

Or put the tissue into the mortar, and add liquid nitrogen to grind fully. Then add the homogenized medium to homogenize.

(2) Mechanical homogenate: Weigh the tissue to EP tube, add the homogenized medium to homogenize the tissue with homogenizer instrument (60 Hz, 90s) in the ice bath. (For samples of skin, muscle and plant tissue, the time of homogenization can be properly prolonged.)

▲ Notes for sample

1. Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
2. If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.
3. If a lysis buffer is used to prepare tissue homogenates, there is a possibility of causing a deviation due to the introduced chemical substance.