

Total Carbonyl Colorimetric Assay Kit

Catalog No: E-BC-K171-M

Method: Colorimetric method

Specification: 96T (Can detect 80 samples without duplication)

Measuring instrument: Microplate reader

Sensitivity: 1.29 $\mu\text{g/mL}$

Detection range: 1.29-45 $\mu\text{g/mL}$

- ▶ Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

General information

▲ Intended use

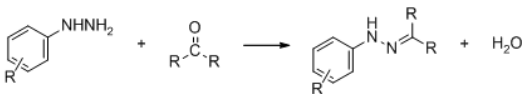
This kit can be used for detection of total carbonyl content in serum, plasma and tissue samples.

▲ Background

Carbonyl is an organic functional group formed by carbon and oxygen. Carbonyl groups (aldehydes and ketones) can be introduced into biomolecules through oxidation. The production of carbonyl groups is considered to be the indirect evidence of the oxidation of biomolecules. The measurement of carbonyl content is helpful to the study of physiology and biochemistry.

▲ Detection principle

Carbonyl can react with 2,4-dinitrophenylhydrazine and produce a kind of reddish brown hydrazone compounds, which has a specific absorbance peak at 370 nm. The content of carbonyl can be calculated according to the absorbance value.



▲ Kit components & storage

| Item | Component | Specification | Storage |
|-----------|----------------------------------|------------------|---------------------------------|
| Reagent 1 | Chromogenic Agent Stock Solution | 1.5 mL × 2 vials | 2-8°C , 6 months, shading light |
| Reagent 2 | 100 µg/mL Standard | 1 mL × 1 vial | 2-8°C , 6 months |
| | Microplate | 96 wells | No requirement |
| | Plate Sealer | 2 pieces | |

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

▲ Materials prepared by users



Instruments

Microplate reader (365-375 nm), Micropipettor, Centrifuge, Incubator, Vortex mixer



Consumptive material

Tips (10 µL, 200 µL, 1000 µL), EP tubes (1.5 mL, 2 mL)



Reagents:

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

▲ Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes.

▲ The key points of the assay

1. The supernatant of sample must be clarified.
2. If the samples are frozen used, centrifuge at 10000 g for 10 min and take the supernatant for measurement.

Pre-assay preparation

▲ Reagent preparation

Preparation of chromogenic agent working solution

Dilute the reagent 1 with double distilled water at the ratio of 1:6. Prepared the fresh solution before use. The prepared solution can be stored at 2-8°C for 7 days.

▲ Sample preparation

The samples should be prepared as conventional methods. Also please refer to appendix II.

▲ Dilution of sample

It is recommended to take 2~3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (1.29-45 µg/mL).

The recommended dilution factor for different samples is as follows (for reference only):

| Sample type | Dilution factor |
|--|-----------------|
| Porcine serum | 1-2 |
| Human serum | 1-2 |
| Human plasma | 1-2 |
| 10% Rat kidney tissue homogenate | 3-5 |
| 10% Rat liver tissue homogenate | 3-5 |
| 10% <i>Epipremnum aureum</i> tissue homogenate | 1 |

Note: The diluent is normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

| Assay protocol | |
|------------------------------|---------|
| Ambient temperature | 25-30°C |
| Optimum detection wavelength | 370nm |

Instructions for the use of transferpettor:

- (1) Equilibrate the pipette tip in that reagent before pipetting each reagent.
- (2) Don't add the liquid outside the tips into the reaction system when pipetting each reagent.

Assay protocol

▲ Plate set up

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|---|---|----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | A | A | S1 | S9 | S17 | S25 | S33 | S41 | S49 | S57 | S65 | S73 |
| B | B | B | S2 | S10 | S18 | S26 | S34 | S42 | S50 | S58 | S66 | S74 |
| C | C | C | S3 | S11 | S19 | S27 | S35 | S43 | S51 | S59 | S67 | S75 |
| D | D | D | S4 | S12 | S20 | S28 | S36 | S44 | S52 | S60 | S68 | S76 |
| E | E | E | S5 | S13 | S21 | S29 | S37 | S45 | S53 | S61 | S69 | S77 |
| F | F | F | S6 | S14 | S22 | S30 | S38 | S46 | S54 | S62 | S70 | S78 |
| G | G | G | S7 | S15 | S23 | S31 | S39 | S47 | S55 | S63 | S71 | S79 |
| H | H | H | S8 | S16 | S24 | S32 | S40 | S48 | S56 | S64 | S72 | S80 |

[Note]: A–H, standard wells; S1–S80, sample wells.

▲ Operating steps

The preparation of standard curve

Dilute 100 µg/mL standard with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 5, 10, 20, 25, 30, 40, 45 µg/mL.

The measurement of samples

- 1) Standard well: add 24 µL of standard with different concentrations into standard wells.
Sample well: add 24 µL of sample into sample wells.
- 2) Add 175 µL of chromogenic agent working solution to each well.
- 3) Mix fully for 5 s with microplate reader and stand for 5 min at room temperature.
- 4) Measure the OD values of each well at 370 nm with microplate reader.

▲ Operation table

| | Standard well | Sample well |
|--|---------------|-------------|
| Standard with different concentrations (µL) | 24 | |
| Sample (µL) | | 24 |
| Chromogenic agent working solution (µL) | 175 | 175 |
| Mix fully for 5 s with microplate reader and stand for 5 min at room temperature. Measure the OD values of each well at 370 nm with microplate reader. | | |

▲ Calculation

Plot the standard curve by using OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the OD value of sample.

The standard curve is: $y = ax + b$. (y: The absolute OD value of standard ($OD_{\text{Standard}} - OD_{\text{Blank}}$); x: The concentration of standard; a: The slope of standard curve; b: The intercept of standard curve.)

1. Tissue homogenates:

$$\text{Total carbonyl content} \quad (\mu\text{g/g}) = (\Delta_{370} - b) \div a \div c$$

2. Serum (plasma):

$$\text{Total carbonyl content} \quad (\mu\text{g/mL}) = (\Delta_{370} - b) \div a \times f$$

Note:

$\Delta A_{370} = OD_{\text{sample}} - OD_{\text{blank}}$ (OD_{blank} is the OD value when the standard concentration is 0).

f: dilution factor of the sample before tested.

c: the content of sample = the wet weight (g) \div the volume of homogenized medium (mL).

▲ Notes

1. This kit is for research use only.
2. Instructions should be followed strictly, changes of operation may result in unreliable results.
3. The validity of kit is 6 months.
4. Do not use components from different batches of kit.

Appendix I Performance characteristics

| Appendix I Performance characteristics | | | |
|--|---------------|----------------------------|-----|
| Detection range | 1.29-45 µg/mL | Average intra-assay CV (%) | 1.9 |
| Sensitivity | 1.29 µg/mL | Average inter-assay CV (%) | 5.0 |
| Average recovery rate (%) | 101 | | |

▲ Example analysis

Take 24 µL of human serum and carry the assay according to the operation table. The results are as follows:

Standard curve: $y = 0.00623x + 0.01228$, the average OD value of the sample is 0.729, the average OD value of the blank is 0.444, and the calculation result is:

$$\text{Total carbonyl content (}\mu\text{g/mL)} = \frac{0.729 - 0.444 - 0.01228}{0.00623} = 43.78 \mu\text{g/mL}$$

Appendix II Sample preparation

The following sample pretreatment methods are for reference only.

▲ Serum

Collect fresh blood and stand at 25°C for 30 min to clot the blood. Then centrifuge at 2000 g for 15 min at 4°C . Take the serum (which is the upper light yellow clarified liquid layer) to preserve it on ice for detection. If not detected on the same day, the serum can be stored at -80°C for a month.

▲ Plasma

Take fresh blood into the tube which has anticoagulant, centrifuge at 700-1000 g for 10 min at 4°C . Take the plasma (which is the upper light yellow clarified liquid layer, don't take white blood cells and platelets in the middle layer) to preserve it on ice for detection. If not detected on the same day, the plasma can be stored at -80°C for a month.

▲ Tissue sample

Take 0.02-1g fresh tissue to wash with PBS (0.01 M, pH 7.4) at 2-8°C . Absorb the water with filter paper and weigh. Homogenize at the ratio of the volume of homogenized medium (2-8°C) (mL): the weight of the tissue (g) =9:1, then centrifuge the tissue homogenate for 10 min at 1500 g at 4°C . Take the supernatant to preserve it on ice for detection. If not detected on the same day, the tissue sample (without homogenization) can be stored at -80°C for a month.

Note:

1. Homogenized medium: Normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4).

2. Homogenized method:

(1) Hand-operated: Weigh the tissue and mince to small pieces (1 mm^3), then put the tissues pieces to glass homogenized tube. Add homogenized medium into homogenized tube, place the tube into the ice bath with left hand, and insert the glass tamping rod vertically into the homogenized tube with the right hand to grind up and down for 6-8 min.

Or put the tissue into the mortar, and add liquid nitrogen to grind fully. Then add the homogenized medium to homogenize.

(2) Mechanical homogenate: Weigh the tissue to EP tube, add the homogenized medium to homogenize the tissue with homogenizer instrument (60 Hz, 90s) in the ice bath. (For samples of skin, muscle and plant tissue, the time of homogenization can be properly prolonged.)

▲ Notes for sample

1. Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
2. If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.