Direct Bilirubin (DBIL) Colorimetric Assay Kit

Catalog No: E-BC-K761-M

Method: Colorimetric method

Specification: 96T (Can detect 46 samples without duplication)

Instrument: Microplate reader

Sensitivity: 0.6 µmol/L

Detection range: 0.6-50 µmol/L

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

General information

▲ Intended use

This kit can be used for detection of direct bilirubin (DBIL) content in serum sample.

▲ Background

Bilirubin is one of the important components of bile. It is the degradation product of hemoglobin in various heme proteins under the action of a series of enzymes. It is important to the digestion and absorption of lipids and the formation of yellow distemper. Bilirubin comes in two forms: water-soluble and water-insoluble. Bilirubin has powerful antioxidant, anti-inflammatory and autoimmune properties. The concentration of bilirubin in human body is related to sex, drug intake, age and so on. Low serum bilirubin is directly related to diabetes, metabolic syndrome, cardiovascular disease and other pathological states. However, high bilirubin is indicative of hemolysis, jaundice, Gilbert syndrome, hepatitis, drug toxicity, and possible bile duct obstruction.

▲ Detection principle

Direct bilirubin react with azo reagent to form azo bilirubin under acidic conditions. The azo bilirubin generated has the maximum absorption at 565 nm. The content of direct bilirubin in serum can be obtained by measuring the change of absorbance.



▲ Kit components & storage

Item	Component	Specification	Storage	
Reagent 1	Acid Agent	30 mL × 1 vial	2-8°C , 6 months, shading light	
Reagent 2	Diazonium Salt	10 mL × 1 vial	2-8°C , 6 months	
Reagent 3	Stop Solution	5 mL × 1 vial	2-8°C , 6 months, shading light	
Reagent 4	Standard	Powder × 2 vials	2-8°C , 6 months, shading light	
N	1icroplate	96 wells		
PI	ate Sealer	2 pieces		

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

▲ Materials prepared by users



Microplate reader (565nm), Micropipettor, Water bath, Incubator, Vortex mixer, Centrifuge

Consumptive material

Tips (10 μ L, 200 μ L, 1000 μ L), EP tubes (1.5 mL, 2 mL, 5 mL)

Reagents

Double distilled water, Normal saline (0.9% NaCl)

▲ Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

▲ Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes

▲ The key points of the assay

- 1. When adding samples, add them quickly or use multiple-channel pipettes.
- 2. There should be no bubbles in the wells of the microplate when measuring the OD value



Pre-assay preparation

▲ Reagent preparation

- 1. Bring all reagents to room temperature before use.
- 2. Preparation of working solution:

Mix reagent 1 and reagent 2 at a ratio of 1.2:1 fully. Prepare the fresh needed amount solution before use

▲ Sample preparation

ice for detection.

- 1. Serum sample: Detect directly
- 2. Sample requirements: There is no hemolysis in the serum sample

▲ Dilution of sample

It is recommended to take 2--3 samples with expected large difference to do pre-experiment before formal experiment and dilute the sample according to the result of the pre-experiment and the detection range (0.6-50 μ mol/LL).

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor		
Human serum	1		
Mouse serum	1		
Rat serum	1		
Rabbit serum	1		
Chicken serum	1		
Porcine serum	1		

Note: The diluent is normal saline (0.9% NaCl) .

Assay protocol					
Ambient temperature	25-30				
Optimum detection wavelength	565 nm				

Instructions for the use of transferpettor:

- (1) Equilibrate the pipette tip in that reagent before pipetting each reagent.
- (2) Don't add the liquid outside the tips into the reaction system when pipetting each reagent.

Assay protocol

▲ Plate set up

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Α	A'	S7	S7'	S15	S15'	S23	S23'	S31	S31'	S39	S39'
В	Α	A'	S8	S8'	S16	S16'	S24	S24'	S32	S32'	S40	S40'
С	S1	S1'	S9	S9'	S17	S17'	S25	S25'	S33	S33'	S41	S41'
D	S2	S2'	S10	S10'	S18	S18'	S26	S26'	S34	S34'	S42	S42'
Е	S3	S3'	S11	S11'	S19	S19'	S27	S27'	S35	S35'	S43	S43'
F	S4	S4'	S12	S12'	S20	S20'	S28	S28'	S36	S36'	S44	S44'
G	S5	S5'	S13	S13'	S21	S21'	S29	S29'	S37	S37'	S45	S45'
Н	S6	S6'	S14	S14'	S22	S22'	S30	S30'	S38	S38'	S46	S46'

Note: A, standard wells; A', standard control wells; S1-S46, sample wells; S1'-S46', sample control wells.



▲ Operation table

- 1. Standard tube: Take 80 µL of reagent 1 into 0.5 mL EP tube. Standard control tube: Take 80 µL of reagent 1 into 0.5 mL EP tube. Sample tube: Take 80 µL of reagent 1 into 0.5 mL EP tube.
 - Sample control tube: Take 80 µL of reagent 1 into 0.5 mL EP tube.
- Add 160 uL of working solution into standard tubes and sample tubes. Add 160 µL of double distilled water into standard control tubes and sample control tubes.
- 3. Add 30 µL of 25 µmol/L standard into standard tubes and standard control tubes
 - Add 30 µL of sample into sample tubes and sample control tubes.
- Mix fully, incubate at 37 for 5 min.
- Add 20 µL of reagent 3 into each tube.
- 6. Mix fully, incubate at 37 for 5 min. Take 250 µL of reaction solution into the corresponding wells and measure the OD values of each well at 565 nm with microplate reader.

▲ Operation table

	Standard tube	Standard _{control}	Sample tube	Sample _{control} tube			
Reagent 1 (µL)	80	80	80	80			
Working solution (μL)	160		160				
Double distilled water (µL)		160		160			
Sample (µL)			30	30			
25 μmol/L Standard (μL)	30	30					
Mix fully and incubate at 37 for 5 min.							
Reagent 3 (µL)	20	20	20	20			

Mix fully, incubate at 37 for 5 min. Take 250 µL of reaction solution into the corresponding wells and measure the OD values of each well at 565 nm with microplate reader

▲ Calculation

DBIL (
$$\mu$$
mol/L) = $A_2 \div A_1 \times C \times f$

Note:

A2: the OD value of sample - the OD value of sample control

A₁: the OD value of standard- the OD value of standard control

C: Concentration of standard (25 µmol/L)

f: Dilution factor of sample before tested

▲ Notes

- 1. This kit is for research use only.
- 2. Instructions should be followed strictly, changes of operation may result in unreliable results.
- 3. The validity of kit is 6 months.
- 4. Do not use components from different batches of kit.



Appendix I Performance characteristics

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Detection range	0.6-50 µmol/L	Average intra-assay CV (%)	4.0			
Sensitivity	0.6 µmol/L	Average inter-assay CV (%)	8.6			
Average recovery rate (%)	100					

▲ Example analysis

Take 30 µL of serum and carry the assay according to the operation table. The results are as follows:

The OD value of the sample is 0.083, the OD value of the sample control is 0.064, the OD value of the standard is 0.150, the OD value of the standard control is 0.041, and the calculation result is:

DBIL content (μ mol/L)=(0.083-0.064)÷(0.150-0.041)×25=4.35 μ mol/L